

Genetic biomarkers in lung cancer therapy – an update

JOANNA CHOROSTOWSKA-WYNIMKO, ADAM SZPECHCINSKI

Laboratory of Molecular Diagnostics and Immunology, National Institute of Tuberculosis and Lung Diseases, Warsaw, Poland

Abstract

Biomarkers are objectively measured indicators of pathogenic processes of disease development or responses to a therapeutic intervention. It is commonly believed that development of reliable biomarkers for lung cancer both prognostic and predictive would considerably change disease diagnostics and treatment outcomes. Paper reviews current data on recent development in clinical applicability of molecular biomarkers in lung cancer.

Key words: biomarker; lung cancer; EGFR.

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From lung carcinogenesis to genetic biomarkers

Technical progress substantially increased our knowledge and understanding of the key role that particular genes play in the pathogenesis of lung cancer. Numerous studies confirmed that the modified expression of genes regulating main biological processes like cell cycle, differentiation, maturation, aging and apoptosis is of decisive significance [1]. It is acknowledged now that unrestrained growth of tumor tissue results directly from the increased activity of oncogenes as well as down-regulated expression of the tumor suppressor genes (TSGs) due to the genetic (mutations) or epigenetic (hyperexpression, methylation) modifications [2]. Since it is possible to effectively detect these alterations in human tissues, some of them might serve as reliable diagnostic markers for cancer screening.

The main mechanisms regulating growth and invasiveness of tumor are reflected in the enhanced or changed expression of particular genes. For example, modified activity of the ERBB gene family encoding EGFR (epidermal growth factor receptor) and HER2/neu (human epidermal growth factor receptor 2) is responsible for the diminished requirement of NSCLC for the growth factors [1, 2]. Similarly, apoptosis that serves as a physiological mechanism regulating cells liveliness, especially in those with disrupted or abnormal DNA

structure, was shown to be inhibited as a result of deregulated expression of p53 and bcl2 genes (respectively in 50% and 30% of NSCLC and more than 90% of SCLC). Other typical alterations of gene expression result in resistance to paracrine growth regulation (loss of heterozygosity (LOH) *TP53*, *p16*), increased angiogenic activity of cancer tissue (VEGF genes), up-regulated tumor cells replication (telomerase gene) as well as augmented ability to invade neighboring tissue and metastasize (laminin and integrin gene) [3].

The number and type of modifications in gene expression parallels cancer development [4]. In addition, certain molecular markers seem characteristic for particular phases of tumor growth and metastasis formation, defining the transition from mild to moderate and severe atypia and subsequently to *carcinoma in situ* (CIS) and microinvasive carcinoma [5].

Early modifications (3p LOH, 9p21 LOH) might be detected as soon as minor lesions such as hyperplasia or dysplasia occur in the bronchial mucosa [6]. Some of them, mostly promoters methylation, have been observed also in normal mucosa of chronic smokers. More significant changes in biomarker expression are found in preneoplastic lesions, in dysplasia or *carcinoma in situ*. Late modifications, typical for invasive cancer, are more abundant and diverse from loss of genetic material (alleles), spontaneous or induced mutations to epigenetic modifications like genes hyperexpression or

methylation. Smoking is particularly effective in inducing multiple genetic modifications in the airways. Active carcinogens present in the cigarette smoke directly interact with the *k-ras*, *p53* and *FHIT* genes critical for the tumor development and induce the earliest carcinogenic modifications – DNA hypermethylation and deletions in the TSGs [7]. Chronic exposition to the cigarette smoke is also responsible for the accumulation of these modifications, increasing therefore the probability of preneoplastic or neoplastic lesions occurrence in the bronchial mucosa. Thus, smoking cessation is rightly regarded as one of the most important methods of lung cancer prevention.

Due to these sequential and progressively expanding changes during the lung cancer development, the expectancies toward genetic biomarkers as the promising diagnostic tool are increasing. Molecular biology techniques might effectively estimate expression of particular, appointed genes not only in tumor cells, but also in other materials like sputum, bronchoalveolar lavage (BAL) and serum/plasma. Number of studies evaluating diagnostic efficiency of different markers or more often marker panels has tremendously increased over the last decade. However, at present there are no widely accepted biomarkers of reliable and confirmed diagnostic value [8, 9].

Clinical implications of genetic markers

Apart from lung cancer screening and diagnostics, molecular markers transpire as a new hope for improved disease prognosis in patients beginning or currently undergoing chemotherapy [10].

Several groups attempted to evaluate a prognostic role of biomarkers in overall survival. However, available data are contradictory and inconclusive. Ramirez et al. observed that *K-ras* mutation in the serum of 12 out of 50 resected NSCLC patients significantly correlated with survival [11]. Also, Kimura et al. found a considerable association between the presence of mutant *K-ras* in plasma and objective responses relevant for the overall survival [12]. Inconsistently, Camps et al. showed no correlation between the presence of mutant *K-ras* genotype in serum and objective response rate, progression-free survival, or overall survival [13]. Moreover, tendency towards the better response rate and survival in patients with circulating mutant *K-ras* was observed. Similarly, it has been demonstrated that structural mutations of *TP53* in the tumor cells, *APC* promoter methylation as well as down-regulated expression of *HIN-1* gene strongly correlated with poor survival of lung cancer patient [14]. Also plasma DNA concentration has been reported to significantly correlate with elevated serum lactate dehydrogenase levels, advanced tumor stage and poor survival in the group of 185 NSCLC [10]. However, practical prognostic impact of above-mentioned biomarkers still remains unclear and needs further evaluation.

Similarly, a reliable serologic biomarker that might be supportive in predicting the treatment response of NSCLC

patients is not available yet. Several molecular diagnostic methods have been evaluated for their applicability in the assessment of lung tumor susceptibility to chemotherapy. Presence of cisplatin adducts in the cytoplasm of normal cells, as well as the decreased expression of *ERCC1* or *Ape1* genes seem to be a reliable and relatively easy method to estimate cancer cells resistance to cytostatic drugs. Likewise, Rosell et al. have proven that high expression of the *RRM1* (ribonucleotide reductase responsible for the DNA synthesis and repair) gene closely corresponded with better outcome of surgical treatment, lower rate of subsequent tumor relapse and much prolonged patients survival time [15]. Another study showed that SNP (single nucleotide polymorphism) in the plasma *MTHFR* (methylene tetrahydrofolate reductase responsible for DNA methylation) is associated with slight differences in median time-to-progression (TTP) in cisplatin/gemcitabine-treated patients with NSCLC [16]. Patients with the *MTHFR 677CC* genotype presented almost two month longer TTP than those with *CT* and *TT* genotypes. Ramirez et al. have demonstrated that methylation of 14-3-3 σ in serum might be a valuable prognostic factor for survival in NSCLC patients receiving platinum-based chemotherapy [17]. The 14-3-3 σ methylation-positive patients (39 out of 115) showed significantly better median survival than patients with sera negative for 14-3-3 σ methylation (15.1 months vs. 9.8 months, respectively). Moreover, the risk of death for 14-3-3 σ methylation-negative responders was almost five times that of 14-3-3 σ methylation-positive responders. Also, circulating nucleosomal DNA in combination with oncological biomarkers: carcinoembryonic antigen (CEA) and *CYFRA21-1* proved its potential value as an early predictor of chemotherapy efficacy [18].

The only molecular test currently validated for the implementation into the clinical practice is the mutational analysis of *EGFR* gene [19]. Epidermal growth factor receptor (*EGFR*) is the molecule of high interest with regard to *EGFR*-targeted treatment with tyrosine kinase inhibitors (TKI) and its certain mutations are established as a marker of patient's response to chemotherapy. Accordingly, Clarke et al. were one of the first to report elevated *EGFR* mRNA in the peripheral blood of 30% NSCLC patients [20]. Next, Kimura et al. detected two major somatic *EGFR* mutations in serum DNA from 13 out of 27 (48.1%) NSCLC patients [21]. *EGFR* mutation positive patients presented better outcomes with gefitinib treatment than *EGFR* mutation negative group. Other authors consistently confirmed that in adenocarcinoma clinically meaningful mutations comprise of base-pair deletion at exon19 (del746_A750) and a point mutation at exon 21 (L858R) [22, 23]. Both result in ligand-independent tumor cell dependence on *EGFR* signaling, therefore enabling *EGFR*-TKI effectiveness. It was shown that these mutations are more frequent in women, of Asiatic origin, non-smokers, diagnosed with adenocarcinoma [24]. Following clinical studies confirmed increased responsiveness, overall survival and tumor free-survival in *EGFR*

mutation positive lung cancer patients. Initially, EGFR TKIs efficacy has been demonstrated in the second-or third line therapy of lung cancer in patients with confirmed EGFR mutations or increased EGFR copy numbers [25, 26]. Recently, Mok et al confirmed that first-line therapy in mutation positive advanced adenocarcinoma patients resulted in better response rate (35.5% vs. 24.4%) and significantly longer progression free survival (29.4 weeks vs 23.4 weeks, $p < 0.0002$) in erlotinib vs. placebo treated group [27]. Similarly, gefitinib assigned as a first-line therapy in treatment naïve, EGFR mutation positive patients with advanced adenocarcinoma allowed significantly prolonged progression-free survival in comparison to standard carboplatin-paclitaxel therapy group [28]. It should be mentioned however, that not all EGFR mutations have similar biological effect. Some, as mutations in exon 20, are related to intrinsic resistance of tumor cells. Moreover, activating mutations of KRAS gene located downstream might cause similar effect of intrinsic resistance to EGFR TKIs. There is also a phenomenon of secondary resistance due to the T790 M specific mutation in EGRF gene or amplification/overexpression of MET gene. Secondary resistance is responsible for TKI-resistant relapse following previously successful treatment with TKIs. Taking above into account it is rather obvious that screening for activating EGFR mRNA or rather DNA mutations predetermining results of treatment with EGFR tyrosine kinase inhibitors will achieve considerable clinical relevance in the near future.

As for the surgical treatment of NSCLC, it is expected that analysis of circulating DNA might prove useful for the post-operative follow-up of NSCLC patients [29]. Monitoring of free plasma DNA concentration has been shown to provide quite valuable information concerning disease recurrence or effectiveness of radical treatment NSCLC. Sozzi et al. confirmed that successful radical tumor resection resulted in significantly lower concentration of plasma DNA than in non-surgically treated patients (7.1 vs. 24.7 ng/ml) [30]. Also, its quantification and molecular characterization was shown to correlate closely with the early recurrence events during the follow-up. In relapse-free individuals circulating DNA concentration was significantly lower than before surgery, while in patients with lung cancer recurrence or metastases up to 20-fold increase was observed together with microsatellite alterations (loci 3p14.2, 3p21, 3p23, 3p24.2, 3p25-26) persistent throughout follow-up period.

Significant correlation between *p16* methylation rate, survival and disease-free survival at 12-month postoperative follow-up has also been reported [31]. The NSCLC patients with *p16* methylation demonstrated in plasma and pre-resection pleural lavage (14.3% and 21.4% of 14, respectively) had shorter survival.

Summary

While discussing the clinical implications of the extensive search for the molecular biomarkers useful in lung

cancer, it should not be forgotten that it is also closely related to the investigation on the new treatment modalities. Many known biomarkers represent key mechanisms required for consecutive stages of tumor development, such as modified requirement for the growth factors or resistance to cell growth and apoptosis regulation. Better understanding of these mechanisms due to the intensive search for the reliable early stage biomarkers might significantly help in elaboration of new treatment concepts.

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References

1. Niklinski J, Hirsch FR (2002): Molecular approaches to lung cancer evaluation. *Lung Cancer* 38: 9-17.
2. Fong KM, Sekido Y, Gazdar AF et al. (2003): Lung cancer 9: Molecular biology of lung cancer: clinical implications. *Thorax* 58: 892-900.
3. Zetter BR (1993): Adhesion molecules in tumor metastasis. *Semin Cancer Biol* 4: 219-229.
4. Pastorino U, Andreola S, Tagliabue E et al. (1997): Immunocytochemical markers in stage I lung cancer: relevance to prognosis. *J Clin Oncol* 15: 2858-2865.
5. Hirsch FR, Franklin WA, Gazdar AF et al. (2001): Early detection of lung cancer: clinical perspectives of recent advances in biology and radiology. *Clin Cancer Res* 7: 5-22.
6. Wistuba II, Behrens C, Virmani AK et al. (1999): Allelic losses at chromosome 8p21-23 are early and frequent events in the pathogenesis of lung cancer. *Cancer Res* 59: 1973-1979.
7. Wistuba II, Gazdar AF, Minna JD (2001): Molecular genetics of small cell lung carcinoma. *Semin Oncol* 28: 3-13.
8. Chorostowska-Wynimko J, Szepechinski A (2007): The impact of genetic markers on the diagnosis of lung cancer: a current perspective. *J Thorac Oncol* 2: 1044-1051.
9. Sonobe M, Tanaka F, Wada H (2004): Lung cancer-related genes in the blood. *Ann Thorac Cardiovasc Surg* 10: 213-217.
10. Gautschi O, Bigosch C, Huegli B et al. (2004): Circulating deoxyribonucleic acid as prognostic marker in non-small-cell lung cancer patients undergoing chemotherapy. *J Clin Oncol* 22: 4157-4164.
11. Ramirez JL, Sarries C, de Castro PL et al. (2003): Methylation patterns and K-ras mutations in tumor and paired serum of resected non-small-cell lung cancer patients. *Cancer Lett* 193: 207-216.
12. Kimura T, Holland WS, Kawaguchi T et al. (2004): Mutant DNA in plasma of lung cancer patients: potential for monitoring response to therapy. *Ann N Y Acad Sci* 1022: 55-60.
13. Camps C, Sirera R, Bremnes R et al. (2005): Is there a prognostic role of K-ras point mutations in the serum of patients with advanced non-small cell lung cancer? *Lung Cancer* 50: 339-346.
14. Marchetti A, Barassi F, Martella C et al (2004): Down regulation of high in normal-1 (HIN-1) is a frequent event in stage I non-small cell lung cancer and correlates with poor clinical outcome. *Clin Cancer Res* 10: 1338-1343.
15. Rosell R, Felip E, Taron M et al. (2004): Gene expression as a predictive marker of outcome in stage IIB-IIIA-IIIB non-

- small cell lung cancer after induction gemcitabine-based chemotherapy followed by resectional surgery. *Clin Cancer Res* 10: 4215-4219.
16. Alberola V, Sarricés C, Rosell R et al. (2004): Effect of the methylenetetrahydrofolate reductase C677T polymorphism on patients with cisplatin/gemcitabine-treated stage IV non-small-cell lung cancer. *Clin Lung Cancer* 5: 360-365.
 17. Ramirez JL, Rosell R, Taron M et al. (2005): 14-3-3 σ methylation in pretreatment serum circulating DNA of cisplatin-plus-gemcitabine-treated advanced non-small-cell lung cancer patients predicts survival: The Spanish Lung Cancer Group. *J Clin Oncol* 23: 9105-9112.
 18. Holdenrieder S, Stieber P, von Pawel J et al. (2004): Circulating nucleosomes predict the response to chemotherapy in patients with advanced non-small cell lung cancer. *Clin Cancer Res* 10: 5981-5987.
 19. Sequist LV, Joshi VA, Janne PA et al. (2006): Epidermal growth factor receptor mutation testing in the care of lung cancer patients. *Clin Cancer Res* 12: 4403-4408.
 20. Clarke LE, Leitzel K, Smith J et al. (2003): Epidermal growth factor receptor mRNA in peripheral blood of patients with pancreatic, lung, and colon carcinomas detected by RT-PCR. *Int J Oncol* 22: 425-430.
 21. Kimura H, Kasahara K, Kawashiri M et al. (2006): Detection of epidermal growth factor receptor mutations in serum as a predictor of the response to gefitinib in patients with non-small-cell lung cancer. *Clin Cancer Res* 12: 3915-3921.
 22. Lynch TJ, Bell DW, Sordella R et al. (2004): Activating mutations in the epidermal growth factor receptor underlying responsiveness of non-small-cell lung cancer to gefitinib. *N Engl J Med* 350: 2129-2139.
 23. Shepherd FA, Rodrigues Pereira J, Ciulena T et al. (2005): Erlotinib in previously treated non-small-cell lung cancer. *N Engl J Med* 353: 123-132.
 24. Gazdar A (2009): Personalized medicine and inhibition of EGFR signaling in lung cancer. *N Engl J Med* 361: 1018-1020.
 25. Rosell R, Moran T, Queralt C et al. (2009): Screening for epidermal growth factor receptor mutations in the lung cancer. *N Engl J Med* 361: 958-967.
 26. Kim ES, Hirsch V, Mok T et al. (2008): Gefitinib versus docetaxel in previously treated non-small-cell lung cancer (INTEREST): randomized phase III trial. *Lancet* 372: 1809-1018.
 27. Mok TS, Wu YL, Chong-Jen Y et al. (2009): Randomized, placebo-controlled, phase II study of sequential erlotinib and chemotherapy as first-line treatment for advanced non-small-cell lung cancer. *J Clin Oncol* 27: 5080-5087.
 28. Mok TS, Wu YL, Thongprasert S et al. (2009): Gefitinib or carboplatin-paclitaxel in pulmonary adenocarcinoma. *N Engl J Med* 361: 947-957.
 29. Rosell R, Fossella F, Milas L (2002): Spanish Lung Cancer Group. Molecular markers and targeted therapy with novel agents: prospects in the treatment of non-small cell lung cancer. *Lung Cancer* 38: 43-49.
 30. Sozzi G, Conte D, Mariani L et al. (2001): Analysis of circulating tumor DNA in plasma at diagnosis and during follow up of lung cancer patients. *Cancer Res* 61: 4675-4678.
 31. Ng CS, Zhang J, Wan S et al. (2002): Tumor p16M is a possible marker of advanced stage in non-small cell lung cancer. *J Surg Oncol* 79: 101-106.